SITE AND NUTRIENT FACTORS CAN INFLUENCE CONIFER RESISTANCE TO INSECT ATTACK

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Managing Insect 'Pests' = Stand Management

- Stand Density
- Stand Composition
- Stand Age



Douglas-fir tussock moth infestation

'Typical' bark beetle infestation

If you are an insect – there is a problem with using plants as food

and two primary questions must be addressed:

• Nutritionally – what do plants (or various plant parts) offer insects?

• What do insects need for optimal growth and reproduction?

Plants can be thought of as <u>a dilute nutrient soup</u> (i.e. amino acids and sugars) in <u>a matrix of structural compounds</u> (such as cellulose and lignin) and <u>allelochemicals</u> (that can include quantitative/qualitative toxins, such as tannins or terpenes respectively).

<u>NITROGEN</u>

- Insects need all of the 'normal' nutrient requirements <u>plus a</u> <u>source of sterols</u>
- Nutrient ratios are not similar across species <u>or plant parts</u>



Xylem tissue is roughly 10X lower in N then is phloem tissue

 Xylem tissue has roughly half the N as foliage

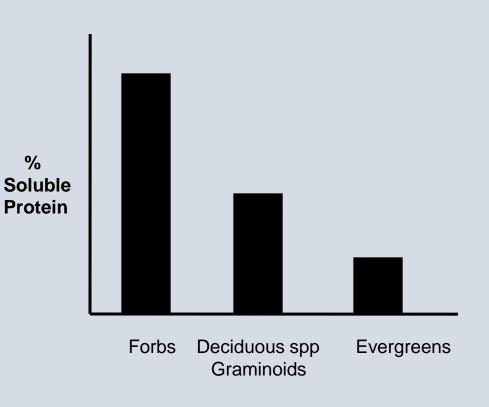
So – how does what an insect eats impact development?

Nitrogen

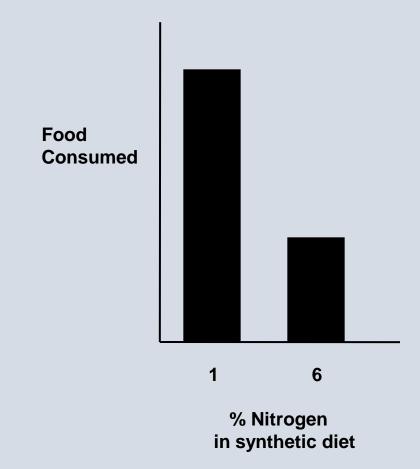
- N is essential to organic life on earth
 - It is a building block for proteins and proteins are the structural material for building insects
 - In insects, proteins > 50% of cuticular dry weight
 - Bulk of plant tissue is comprised of <u>carbohydrates</u> (cellulose, lignin, etc)
- Although it is a common chemical in the atmosphere N is not very available in a usable form
- N is also frequently combined with other elements, making it more difficult for plants/insects to capture and utilize

Nitrogen

- Growth efficiency of insects if often correlated with protein content of their food
- N in plants varies by species, organ, season and environmental factors
- Plant amino acids differ from what is required for insect growth, development and reproduction

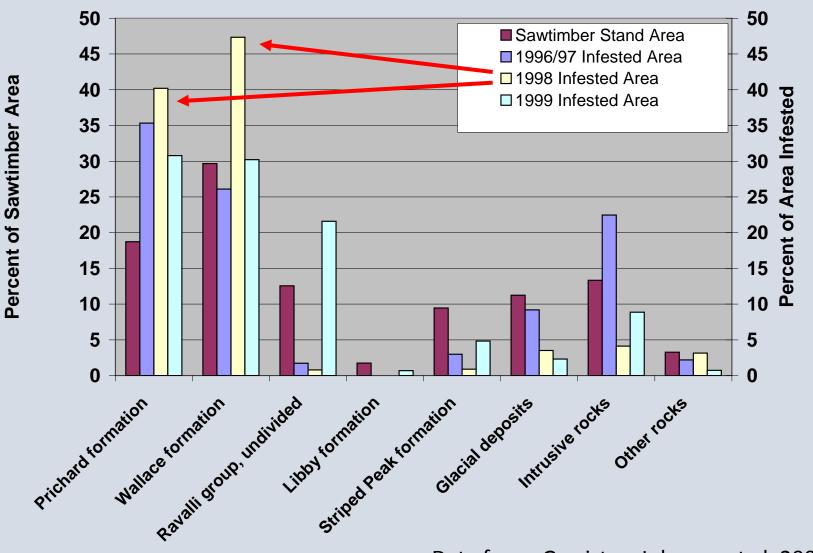


Nitrogen



- However, higher total N in plants may not coincide with usable N for insects because several classes of plant defensive chemistry have N as a building block (i.e. alkaloids and tannins)
- These compounds can/do reduce plant digestibility

Can nutrient measurements be used in predicting stand susceptibility?

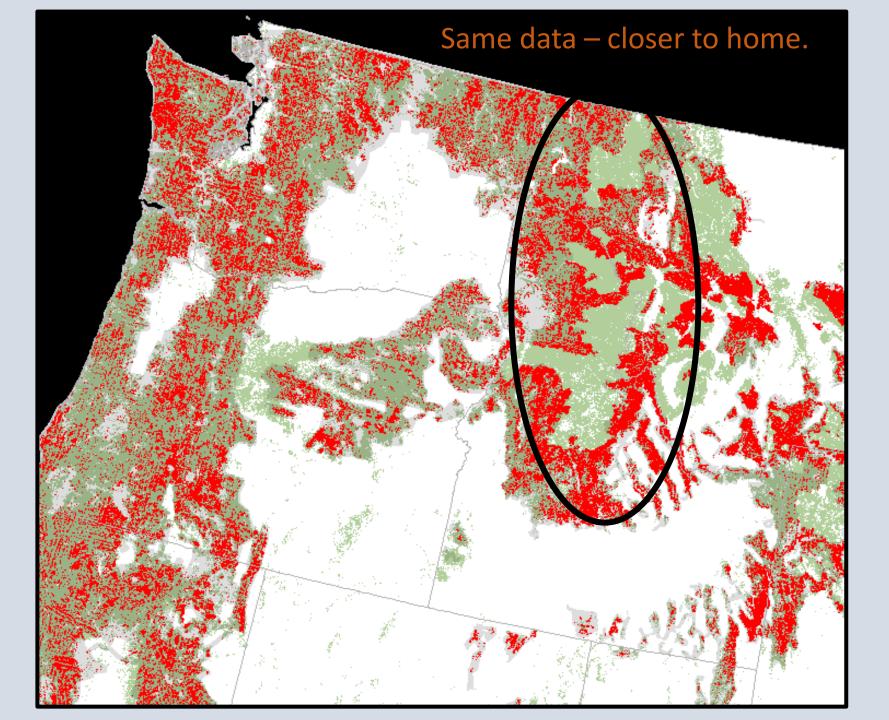


Good Rock – Bad Rock

- Why is rock type important?
 - Different nutrient concentrations.
- Metasedimentary rocks are poor nutrient producers.
 - Nitrogen, Potassium, Sulfur and Boron

Data from: Garriston-Johnson et al. 2003

Forest tree <u>mortality</u> from insect herbivores. Represented by the red pixels. From: USDA Forest Service



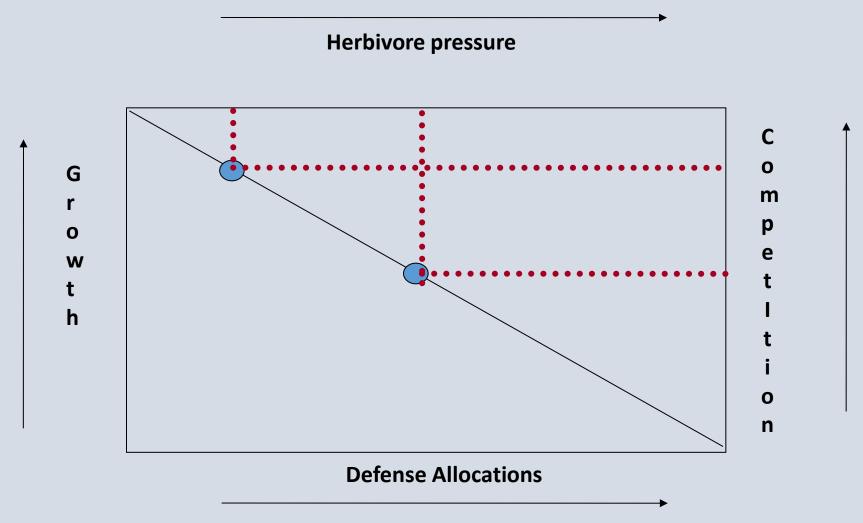
In other words, the obvious question became –

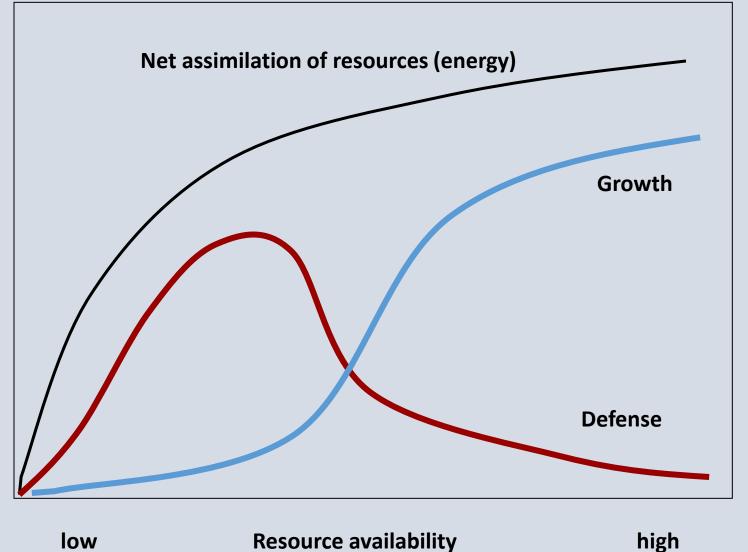
Why the bulls-eye on Idaho?

One Hypothesis Based upon established plant-insect interaction theories.

- There is a continuum within plants of resource allocation between growth and differentiation
- One end of the continuum (<u>in resource rich environments</u>) has plants being selected based upon competition (growth characteristics)
- The other end of the continuum (<u>in resource poor environments</u>) suggests plants are selected based upon herbivore defense (differentiation characteristics)
- Thus the dilemma faced by plants: Grow fast enough to compete with other plants while defending themselves against herbivores and pathogens

Competition and herbivory together will select for certain levels of defense allocation, if there is a cost to defense there is a tradeoff between these strategies





A plant has a finite amount of energy that is allocated among pools used for growth, reproduction and defense.

In this scenario – growth includes reproduction.

Energy is split between growth activities and 'defense' activities.

Mountain Pine Beetle, Dendroctonus ponderosae





Can we modify the relationship between the beetle, its host and its associated fungi by altering some basic tree chemistry?

Where does the developing larva get its N?

How involved are the fungi with larval nutrition?

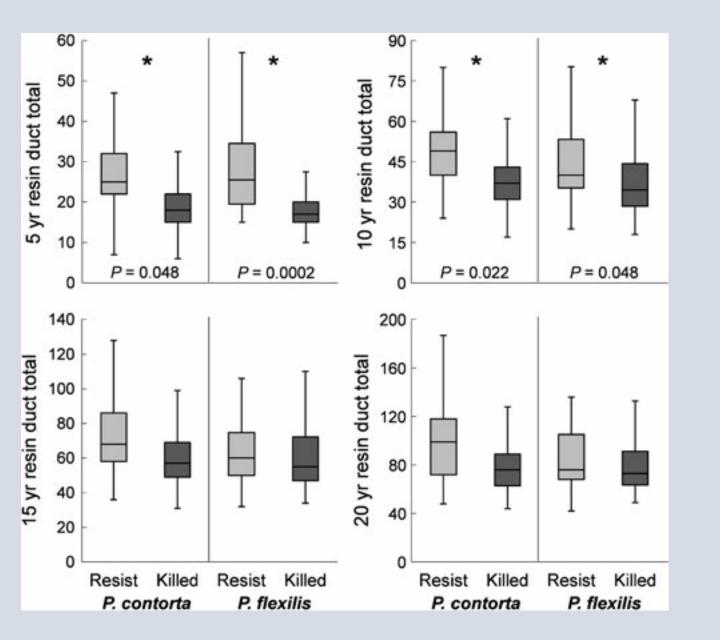
Does tree nutrition play a role?

Review of tree resistance mechanisms



Constitutive Defenses = Resin Flow To some extent, always present Induced Response = Hypersensitive Lesion Occurs following attack (beetle or fungus)





Tree Resistance Mechanisms (continued – but new).

Vertical Resin Canal Comparison:

Lodgepole and Limber pines that resisted or succumbed to attack by MPB.

From: Ferrenberg et al. 2013. Oecolgia (online)

MPB-Lodgepole-Fungal Interactions

- Two field sites:
 - Craig Mountain
 - University of Idaho Experimental Forest
- Fertilizer applied to individual trees (fall or winter)
 - Measure inner bark N content
 - Measure resin flow
 - Measure inner bark monoterpene content (in progress)
- Controlled laboratory studies focused on:
 - Grosmannia clavigera
 - Ophiostoma montium

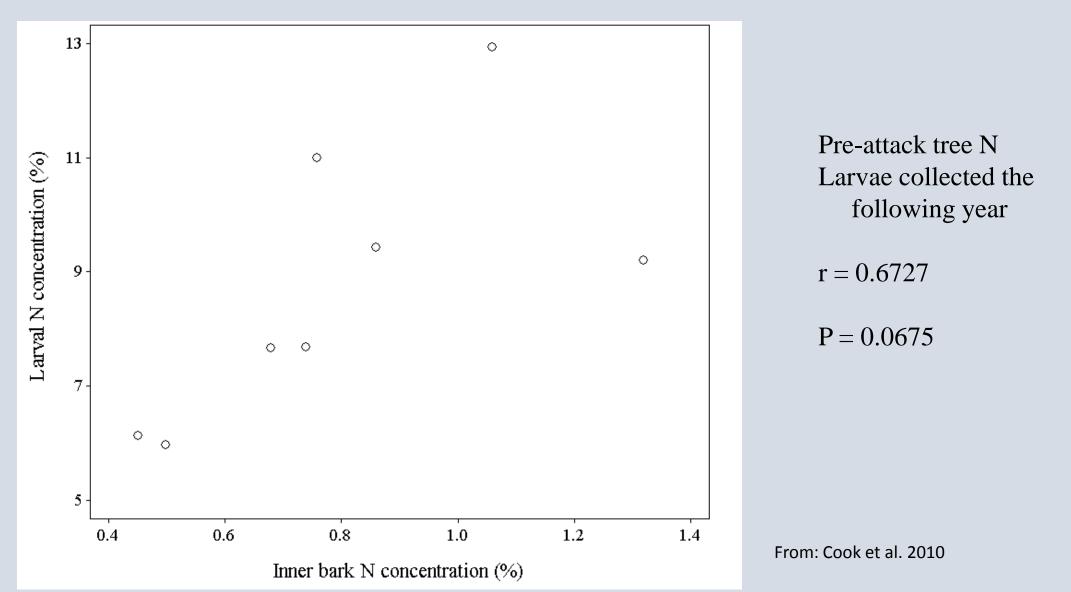
What happens when you apply fertilizer: Fertilizer applied in March, Measurements in July Change in inner bark N content (dry weight)

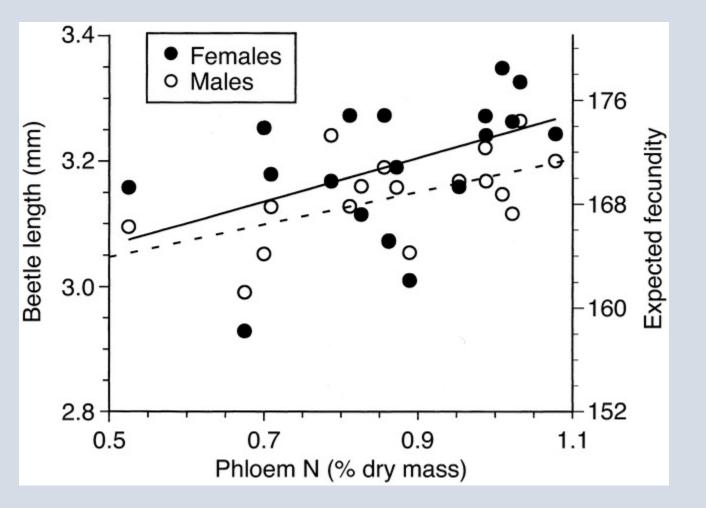
N treatment	Pre-fertilization	Post- fertilization	Difference
Control, 0 lbs/ac	0.50 <u>+</u> 0.03 a	0.51 ± 0.02	0.04 <u>+</u> 0.03 a
Low, 300 lbs/ac	0.52 ± 0.02 a	0.78 ± 0.07	0.31 ± 0.07 b
High, 600 lbs/ac	0.55 <u>+</u> 0.03 a	0.75 ± 0.08	0.19 <u>+</u> 0.11 a

From: Cook et al. 2010

Methods and Results

Correlation between tree inner bark and larval N contents





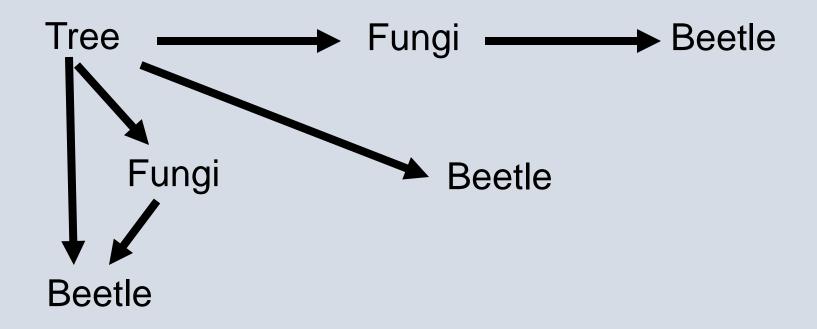
Tree N and size of SPB

From: Ayres et al. 2000 Ecology

Beetle size is usual correlated with survival, dispersal and fecundity

Bigger is better for the beetle

Trophic movement of N:

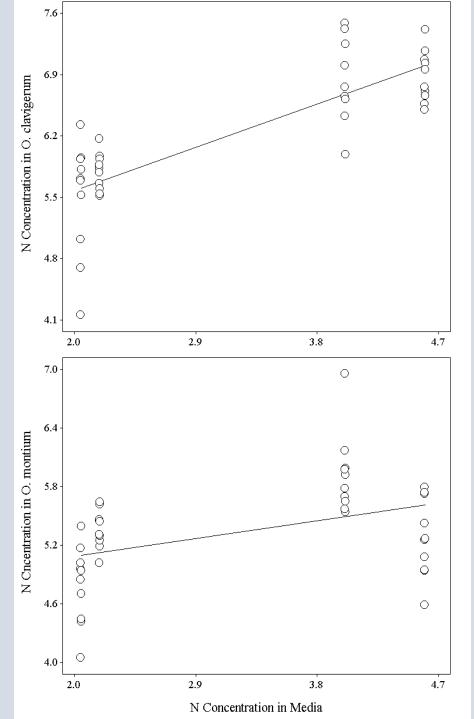


There will be a different shift in isotopic N content depending on the source of the N acquired by the developing beetle. Strong linear relationship between N concentration in the growth media and *G. clavigera*

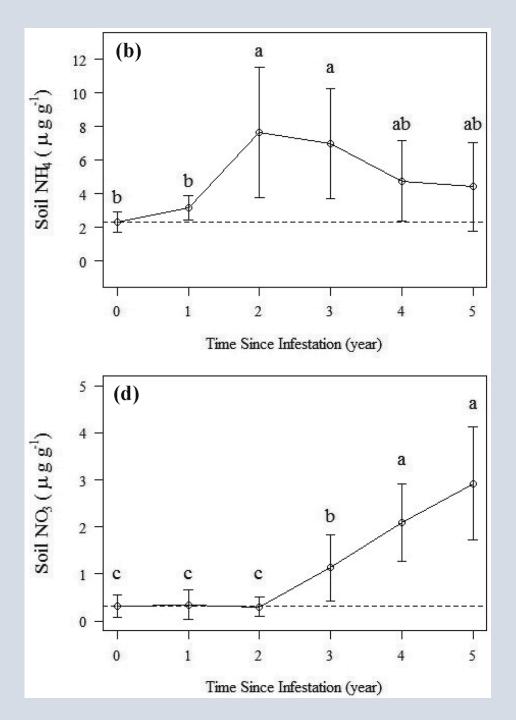
R2 = 0.8116

Weaker relationship when we Examine the *O. montium*

R2 = 0.1801



From: Cook et al. 2010

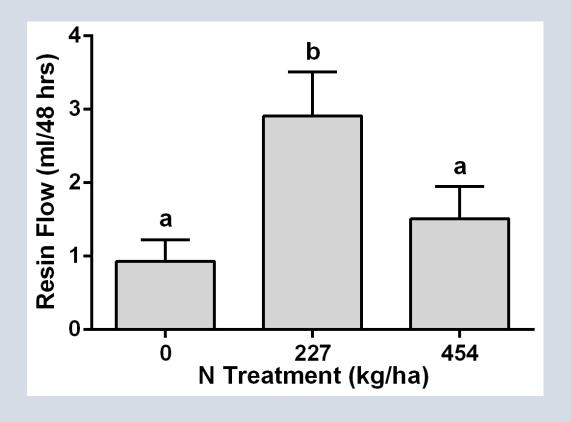


Soil nitrogen five years after bark beetle infestation in lodgepole pine forests

From: Norton et al. 2015. Soil Science Society of America Journal (online)

Nitrogen fertilization of individual Trees at 3 concentrations (0, 200 and 400 lbs/ac)

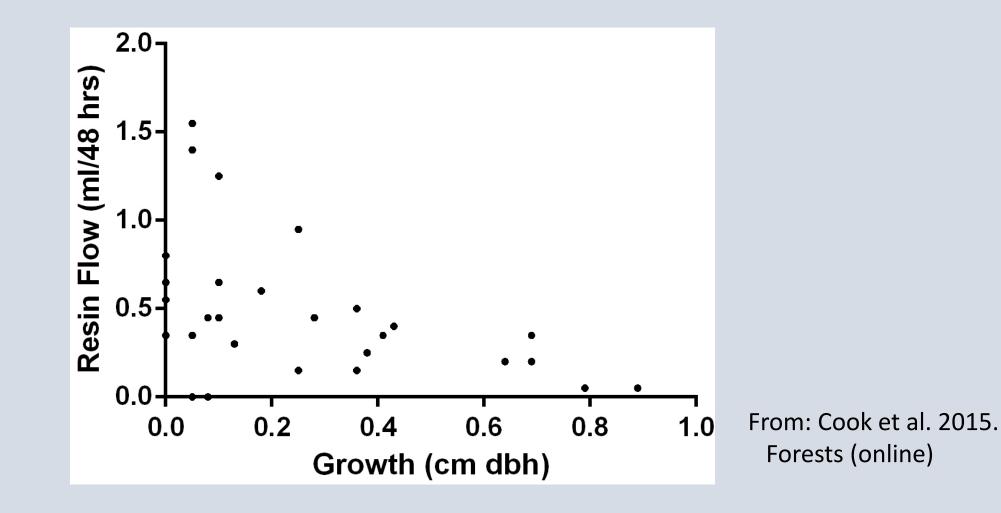
Fertilizer applied October 2007 Resin flow measured in July, 2008



One question to ask – how much is too much?

From: Cook et al. 2015. Forests (online)

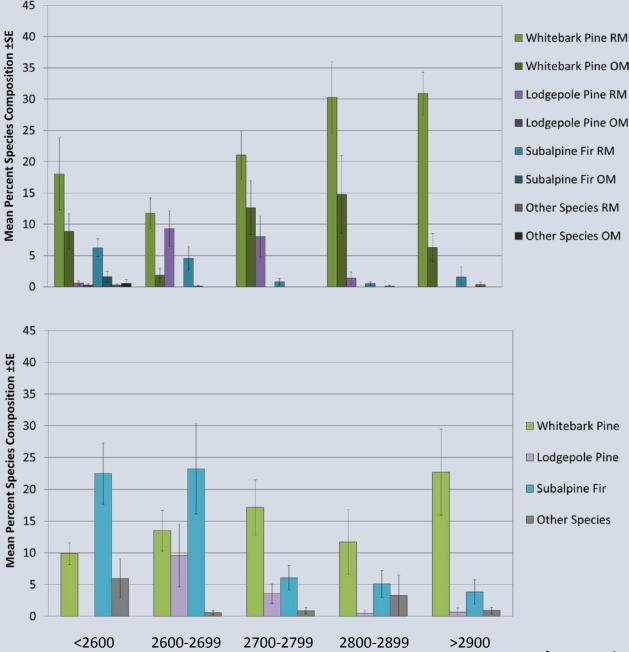
Lodgepole Pine: Strong negative relationship between Growth and Resistance Mechanisms



In many ways Mountain Pine Beetle infestations in Whitebark Pine Provide a 'slower-moving' system in which to examine the interactions.



Photo from: Kendra Schotzko

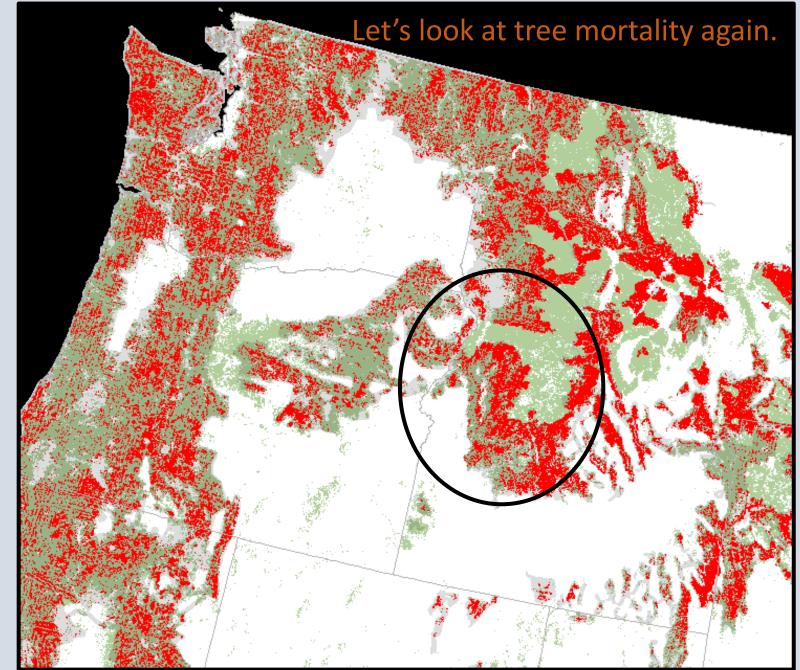


Stand Compositions are modified following infestations by Mountain Pine Beetle – but at most elevations: there are residual whitebark and lodgepole pines that remain.

Several factors are at play including: climate differences blister rust intensity competition others.

We have initiated a project that also compares chemistry of the residual trees – University of Idaho collaboration with University of Wisconsin and University of Alberta.

Data from: Kendra Schotzko

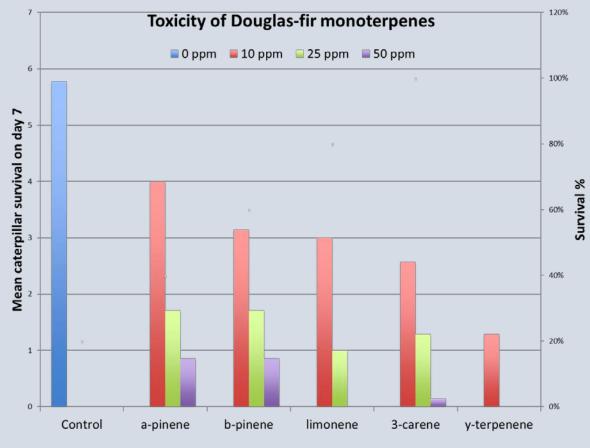


Mountain Pine Beetle Other Bark Beetles Balsam Woolly Adelgid Spruce Budworm Douglas-fir Tussock Moth Other Pests

Fertilization and Foliar Chemistry – Impact on Defoliators Douglas-fir Tussock Moth

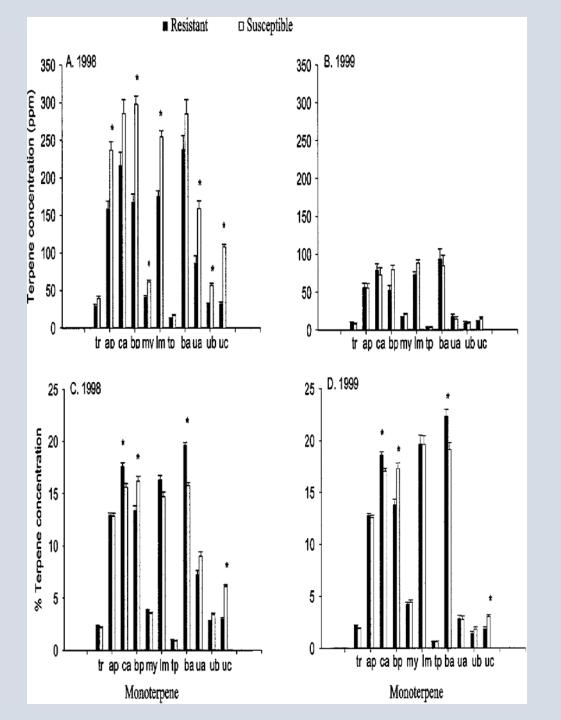


Treatment N applied	Total monoterpenes (mg/g foliar tissue)		
Control (0)	7.0 <u>+</u> 0.9 a		
Low (227 kg/ha)	8.6 <u>+</u> 1.2 a		
High (454 kg/ha)	6.9 <u>+</u> 0.6 a		



Douglas-fir monoterpenes (at three different concentrations)

Data from: A. Carroll



Western Spruce Budworm

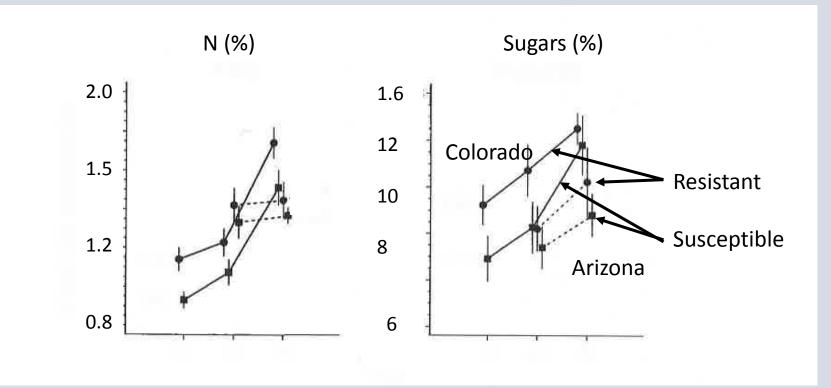
Prior work indicates that there were significant differences in the concentration and/or percentage of some individual monoterpenes present in resistant versus susceptible Douglas-fir.

This prior work also showed a decrease in potential fitness of the insect as overall monoterpene concentrations increased.

From: Chen et al. 2002

Western Spruce Budworm

Data from: Clancy et al. 1993



There were also differences in the %age of N and Sugars between resistant and susceptible Douglas-fir foliage at two sample locations (Colorado and Arizona)

Fertilization and Foliar Chemistry – Impact on Defoliators Douglas-fir Tussock Moth

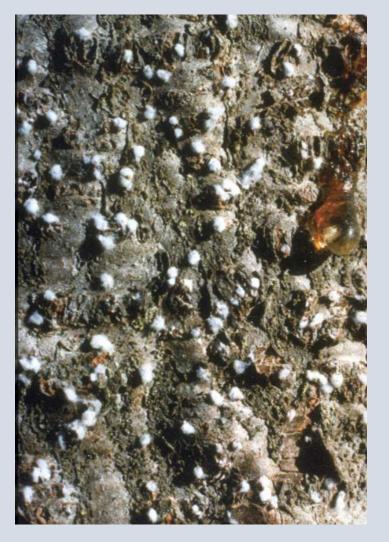
Treatment N applied	N	В	Р	К	S
Control 0 kg/ha	1.2 <u>+</u> 0.1	24.2 <u>+</u> 2.1	0.2 <u>+</u> 0.01	0.7 <u>+</u> 0.04	0.1 <u>+</u> 0.01
Low 227 kg/ha	1.4 <u>+</u> 0.2	21.8 <u>+</u> 4.0	0.2 <u>+</u> 0.01	0.6 <u>+</u> 0.05	0.1 <u>+</u> 0.01
High 454 kg/ha	1.4 <u>+</u> 0.1	23.3 <u>+</u> 3.7	0.2 <u>+</u> 0.01	0.7 <u>+</u> 0.04	0.1 <u>+</u> 0.01

Data from: A. Carroll

- Other elements were also examined
- No change in foliar chemistry based upon fertilization (measured too early)
- Few correlations between individual monoterpenes and individual elements
- Need to refine tests
- Correlations between individual elements and Spruce Budworm performance have been reported

Summary

- We can modify tree chemistry in such a way as to impact tree resistance and insect survival.
- By modifying tree chemistry, we can also impact the physical parameters of a tree that are important as resistance parameters.
- Individual treatments need to be assessed in relation to multiple factors including soil type, current pest status, management objectives, etc.
- There is still a lot of work to do and quite a few 'new' challenges.



Balsam Woolly Adelgid on Subalpine Fir

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If I left any time – Questions anyone?

